

(+)-Episesaminone, a *Sesamum indicum* Furofuran Lignan. Isolation and Hemisynthesis

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Several lignans from *Sesamum* sp. contain an unusual oxygen insertion between their furano and aromatic rings. As part of our ongoing studies to clarify the biosynthetic pathway to the sesame lignans, the furanoketone, (+)-episesaminone, was isolated and fully characterized in part via hemisynthesis from (+)-sesamol.

During our ongoing studies directed toward defining how (+)-sesamol (**1**) biosynthesis occurs in *Sesamum indicum* L. (Pedaliaceae),¹ as well as that of related lignans,^{2–6} the hitherto unknown tetrahydrofuran lignan, (+)-episesaminone (**2**) (Figure 1), was isolated from commercially available unroasted and unbleached seeds, as well as from freshly harvested seed tissue. Its structure was established via a combination of ¹H–¹³C NMR, IR, UV, and MS analyses and confirmed by total synthesis as follows: the presence of a conjugated ketone in (+)-episesaminone (**2**) was suggested from its IR (1647 cm⁻¹) and UV (312 nm) absorbances and was further supported from the resonance at 197.3 ppm in its ¹³C-NMR spectrum, which revealed an aromatic conjugated carbonyl group, with the latter confirmed by use of a ¹H–¹³C 2D NMR HMBC experiment. The ¹H-NMR spectrum showed two aromatic *o*-carbonyl protons at 7.56 ppm (1H, dd, *J* = 8.1 Hz, *J* = 1.7 Hz) and 7.45 ppm (1H, d, *J* = 1.7 Hz). Additionally, the proton resonances at 7.56 and 7.45 ppm, together with those at 6.86 ppm (1H, d, *J* = 8.1 Hz) and two methylenedioxy protons downfield at 6.05 ppm, were indicative of a 3,4-methylenedioxyphenyl (piperonyl) acetophenone system. This was additionally supported by its mass spectrum, which exhibited a base peak at *m/z* 149 and a fragment ion at *m/z* 121, corresponding to acylium ion [ArCO⁺] and piperonyl ion [Ar⁺] fragments, generated after carbon–carbon bond cleavage at C-7'/C-8' and C-7/C-1', respectively.⁷ In an analogous manner, the second piperonyl system was established by the proton resonances at 6.95, 6.84, 6.76 (aromatic protons), and 5.94 ppm (OCH₂O). This was further confirmed from its ¹³C-NMR spectrum, using both DEPT and HMBC, to determine the multiplicities and ³*J* coupling relationships. That is, the methylenedioxy bridges at 5.94 and 6.05 ppm in the ¹H-NMR spectrum were correlated with the two OCH₂O groups at 101.13 and 102.01 ppm, respectively, thereby confirming the piperonyl structure for both aromatic substituents in the ¹³C–¹H HETCOR NMR spectrum. The remaining assignments were made using a combination of ¹H, ¹H–¹H COSY, ¹³C (DEPT mode), and ¹³C–¹H HETCOR NMR spectroscopic analyses, which established the tetrahydrofuran skeleton as follows: the doublet at 4.64 ppm (*J* = 9.1 Hz)

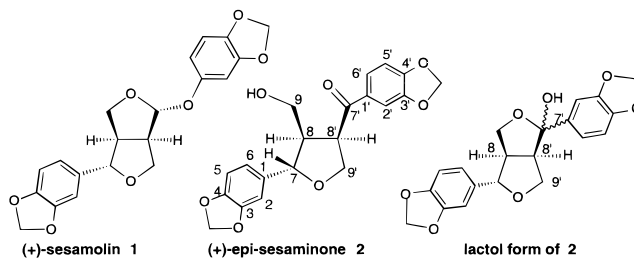


Figure 1. Lignans from *Sesamum indicum*.

was assigned to the oxybenzylic proton H-7 in a trans configuration to H-8. Next, the multiplets centered at 4.28 and 4.10 ppm were correlated in the ¹H–¹H COSY NMR spectrum and assigned to the three proton resonances, viz. the protons of the methylene group at C-9' and the methine proton at C-8'. In the same way, the multiplets at 3.76–3.66 and 2.86 ppm were correlated in the 2D ¹H–¹H spectrum and assigned to the hydroxymethyl group at C-9 and the methine proton at C-8. Furthermore, 2D ¹³C–¹H HETCOR NMR spectroscopic studies confirmed the furanoketone structure by assignment of chemical shifts in the ¹³C-NMR spectrum relative to the corresponding ¹H-NMR resonances (see Experimental Section), in a manner analogous to that used for other naturally occurring keto-lignans.^{8,9} Thus, the methylenic multiplets at 4.28–4.10 ppm and 3.76–3.66 ppm displayed correlations with the methylenic carbon at 70.84 (C-9') and the CH₂OH group at 61.29 ppm (C-9), respectively, as did the multiplets at 4.18 and 2.86 ppm with the methine carbons at 49.98 ppm (C-8') and 52.26 ppm (C-8) in the DEPT ¹³C-NMR spectrum. Additionally, the H-7 proton at 4.64 ppm was correlated with the methine carbon (C-7) at 83.70 ppm, thereby accounting for all of the resonances of the tetrahydrofuran skeleton. Further support for this structural determination was made by MS analysis, which displayed a molecular ion at *m/z* 370, together with fragment ions at *m/z* 203, 149, and 121 corresponding to [ArC₅H₆O⁺], [ArCO⁺], and [Ar⁺], respectively.⁷

Next, nuclear Overhauser enhancement difference experiments established a cis stereochemistry between the H-8 and H-8' protons on the bridgehead carbons. Furthermore, since no NOE was observed between the H-7 and H-8 protons, these therefore had a trans stereochemical relationship between them. Thus, (+)-episesaminone (**2**) has the same overall stereochemistry as that of magnolenin C (**3**) from *Magnolia grandiflora*¹⁰ (Figure 2). The absolute configuration of **2** at C-8' was

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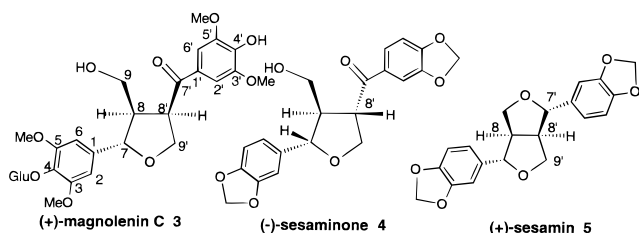


Figure 2. Lignan metabolites isolated from *Magnolia grandiflora* and *Sesamum indicum*, as well as from *Streptomyces* cell cultures grown on sesame-seed media.

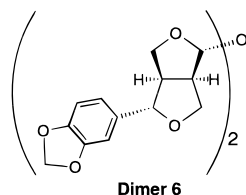
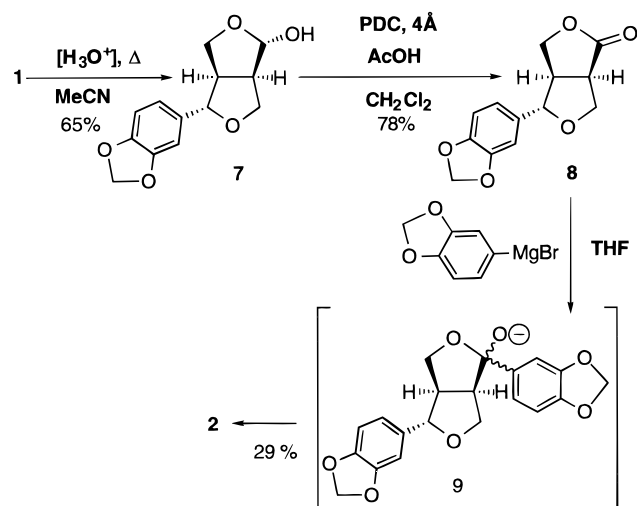


Figure 3. Dimer **6** obtained by acid hydrolysis of sesamol **1**.

Scheme 1. Hemisynthesis of (+)-Episesaminone (**2**) from (+)-Sesamol (**1**)



deduced as *R* ($[\alpha]_D + 126.2^\circ$; c 0.127, CHCl_3), given that the previously reported *S* epimer,¹¹ (–)-sesaminone (**4**) had a negative optical rotation ($[\alpha]_D - 25.0^\circ$; c 0.14, MeOH).

To establish unambiguously the proposed structure of (+)-episesaminone (**2**), its hemisynthesis from (+)-sesamol (**1**),¹² a readily available sesame-seed lignan together with (+)-sesamin (**5**),¹³ was next developed. The approach used was based on the known chemistry of (+)-sesamol (**1**), which can be hydrolyzed in H_2SO_4 – EtOH to give, as the major products, the dimer **6** (Figure 3) and lactol (+)-samin (**7**).¹⁴ Subsequent modification of this hydrolytic procedure with **1** in acetate buffer (165 mM, pH 1.0) in EtOH gave three compounds: the symmetrical dimer **6** (disaminyl ether) as the major product, as well as smaller yields (10–20%) of the required (+)-samin (**7**) and its corresponding ethoxyether.¹⁵

Further refinement of the hydrolytic procedure, however, using DOWEX-50 X 2–200 exchange resin, NaOAc buffer (165 mM, pH 1.0), and 10% HCl in CH_3CN gave **7** in 65% yield (Scheme 1).

From lactol **7**, the key step to afford (+)-episesaminone (**2**) was a mono-Grignard addition to lactone **8** as

shown in Scheme 1. Thus, oxidation of (+)-samin **7** with pyridinium dichromate (PDC) under acidic conditions^{1,16} afforded the corresponding lactone, (+)-acuminatolide **8**^{1,17} in 78% yield. Subsequent condensation of **8** with the Grignard reagent,¹⁸ obtained from bromosessamol (1-bromo-3,4-(methylenedioxy)benzene), at room temperature, followed by hydrolysis of the intermediate lactol **9**, gave (+)-episesaminone (**2**) as shown (Scheme 1). The synthetic product was identical in all respects to naturally occurring (+)-episesaminone (**2**) (see Experimental Section), except for the optical rotation value, i.e., $[\alpha]_D + 120^\circ$ (c 0.127, CHCl_3) versus that of the natural product **2** [$[\alpha]_D + 126.2^\circ$ (c 0.127, CHCl_3)]. Thus, since (+)-samin **7** has a *R* configuration at C-8',^{1,14} this confirms the proposed stereochemical assignment of (+)-episesaminone (**2**).

Last, in terms of the biosynthesis of (+)-episesaminone (**2**), it can tentatively be proposed that it is formed in vivo from (+)-sesamin (**5**) (Figure 2). Given that (+)-episesaminone (**2**) can exist in its lactol form (see Figure 1) under basic conditions,¹⁰ only a single oxidation step from (+)-sesamin **5** to **2** is required. Additionally, it should be noted that its C-8' *S* epimer, (–)-sesaminone (**4**) (Figure 2) has been reported as present in *Streptomyces* IT-44 cell cultures grown on sesame-seed media.¹¹ A possible explanation for its formation is that cis (+)-episesaminone (**2**) is epimerized at C-8' under prolonged basic treatment to give the more stable trans form **4**.

Experimental Section

Plant Materials. Seeds of *S. indicum* (unroasted and unbleached) were either purchased from a local produce store or obtained from ripe pods of mature *S. indicum* plants grown in greenhouses at Washington State University.

General Experimental Procedure. Solvents were either HPLC grade (CH_3CN , MeOH , AcOH , CH_2Cl_2 , THF) or ACS grade (EtOAc , Hexanes) from Baker. Tetrahydrofuran (THF) and methylene chloride (CH_2Cl_2) were distilled over LiAlH_4 –triphenylmethane and CaH_2 , respectively. Column and analytical TLC separations (Al Si G/UV 254) were performed using Si gel 60 (230–400 mesh) (Whatman). IR spectra were recorded on a Perkin-Elmer 1720-X FTIR spectrometer, whereas NMR spectra were obtained using either Bruker AMX300 (for ^1H , ^{13}C , and 2D NMR) or Varian 500VXR (for NOE measurements) spectrometers. All spectra were recorded in CDCl_3 using TMS as internal reference, with chemical shifts (δ) expressed in parts per million and coupling constants (J) in Hertz. UV and ORD spectra were recorded on a Perkin-Elmer Lambda 6 UV/vis spectrophotometer and a JASCO 181 polarimeter at λ 289 nm (Na), respectively. HPLC separations were carried out in the reversed-phase mode [Waters C_{18} Nova-Pak, 150×3.9 mm i.d., $4 \mu\text{m}$], eluted with MeOH – H_2O in a linear gradient of 3:7 \rightarrow 7:3 over 60 min with detection at 280 nm. EIMS analyses were carried out with an HPLC–MS (Integrity Waters), using a reversed-phase column (Waters C_{18} Nova-Pak, 150×2 mm i.d., $4 \mu\text{m}$) eluted with a linear solvent gradient A: H_2O (3% v/v acetic acid) and B: CH_3CN ; as follows: 8:2 \rightarrow 7:3 in 10 min, then 7:3 \rightarrow 1:1 in 25 min; MS analyses TIC (Total Intensity Chromatogram) were performed from m/z 80 to 700 at a rate of 1 scan s^{-1} , with an optimal temperature of the nebulizer at 80°C

using (+)-sesamin (**5**) as a reference. Interpretation of fragmentation patterns were calculated where Ar = 3,4-methylenedioxyphenyl.⁷ HRMS analyses employed a VG 7070 EHF mass spectrometer at 70 eV.

Isolation of (+)-Episesaminone (2). Commercially available *S. indicum* seeds (1.28 kg, unroasted) were homogenized in a Waring blender containing MeOH (1 L) and extracted exhaustively with hot MeOH (11 × 1 L) at 50 °C. The resulting MeOH solubles (12 L) were combined, cooled to 0 °C for 45 min, then filtered with the filtrate concentrated in vacuo to give an oily residue. This residue was suspended in MeOH–H₂O (1:1, 150 mL), then sequentially extracted with hexanes (2 × 400 mL) and EtOAc (4 × 500 mL). The EtOAc solubles were combined and concentrated (15 mL), to which MeOH (150 mL) was next added. The resulting precipitate (3.24 g) was removed by filtration to give a crude mixture of (+)-sesamin (**5**) and (+)-sesamolol (**1**),^{9,19} with the filtrate concentrated to afford a yellow gum (12.3 g). The latter was dissolved in a minimal amount of EtOAc and applied to a Si gel column (300 g) eluted with a hexane–EtOAc gradient (5:1 to 1:1; 150 mL each). Fractions 6–7 (25 mL each) and 8–9 contained (+)-sesamin (**5**) (536 mg)¹³ and (+)-sesamolol (**1**) (279 mg),⁹ respectively. Fractions 13–15 were evaporated to dryness, reconstituted in CHCl₃, then subjected to preparative TLC eluted with 3% (v/v) MeOH in CHCl₃ to give (+)-episesamin (2 mg) (*C*-7' epimer of **5**),⁷ (+)-kobusin (1.5 mg),²⁰ and a more polar fraction. The latter was subjected to analytical HPLC separation [C₁₈ reversed-phase, eluted with MeOH–H₂O, linear gradient 3:7 → 7:3 in 60 min] to give (+)-piperitol (2 mg)²¹ and (+)-sesamolol (1 mg).²² Fractions 17–20 (58 mg) were pooled and purified by preparative TLC (CHCl₃–MeOH 5:1) to give two fractions, purification of which by analytical HPLC as before gave (+)-sesamolol (1.5 mg)²² and (+)-episesaminone (**2**) (1 mg), respectively. Fractions 21–24 (88 mg) were chromatographed on Si gel eluted with 5% (v/v) MeOH in CHCl₃ to afford (+)-pinoselinol (23 mg)²³ and *trans*-ferulic acid (7 mg).

(+)-Episesaminone (2). (+)-[3*R*-(3 α ,4 α ,5 β)]-1,3-Benzodioxol-5-yl[1-(1,3-benzodioxol-5-yl)-2-(hydroxymethyl)tetrahydro-1*H*,3*H*-3-furanyl]methanone: IR (CHCl₃) ν_{\max} 3600–3200 (OH), 1647 (C=O), 1604, 1505, 1444, 1409 (C=C Ar), 1251 (C–O) cm⁻¹; UV (MeOH) λ_{\max} (log ϵ) 230 (4.76), 280 (3.75), 312 (2.87) nm; [α]_D +126.2° (*c* 0.127, CHCl₃); MS *m/z* 370 [M⁺] (38), 352 [M – H₂O⁺] (5), 243 (5), 203 [ArC₅H₆O⁺] (25), 194 (40), 176 (48), 152 (50), 150 [ArCHO⁺] (45), 149 [ArCO⁺] (100), 121 [Ar⁺] (40); HRMS *m/z* 370.1044 (calcd for C₂₀H₁₈O₇, 370.1053); ¹H-NMR δ 7.56 (1H, dd, *J* = 8.1 Hz, *J* = 1.7 Hz, H-6'), 7.45 (1H, d, *J* = 1.7 Hz, H-2'); 6.95 (1H, d, *J* = 1.65 Hz, H-2), 6.86 (1H, d, *J* = 8.1 Hz, H-5'), 6.84 (1H, dd, *J* = 7.94 Hz, *J* = 1.65 Hz, H-6), 6.76 (1H, d, *J* = 7.94 Hz, H-5), 6.05 (2H, s, OCH₂–O), 5.94 (2H, s, OCH₂–O), 4.64 (1H, d, *J* = 9.1 Hz, H-7), 4.28 (1H, m, H-9'eq), 4.10 (2H, m, H-9'ax + H-8'), 3.76 (1H, m) and 3.66 (1H, m, H-9), 2.86 (1H, m, H-8); ¹³C-NMR (DEPT mode) δ 197.30 (s, C-7'), 152.18 (s, C-4'), 148.44 (s, C-3'), 147.98 (s, C-4), 147.45 (s, C-3), 134.42 (s, C-1), 131.14 (s, C-1'), 124.93 (d, C-6'), 119.08 (d, C-6), 108.30 (d, C-5), 108.09 (d, C-2'), 107.93 (d, C-5'), 107.14 (d, C-2), 102.01 (t, OCH₂O), 101.13 (t, OCH₂O), 83.70 (d, C-7), 70.84 (t, C-9'), 61.29 (t, C-9), 52.26 (d, C-8), 49.98 (d, C-8') ppm.

Ripe Pod Extraction. Mature (9–12 weeks' growth) *S. indicum* greenhouse grown plants were harvested, with the intact seeds (50 g) carefully removed, and immediately homogenized in a Waring blender containing MeOH (200 mL) and then extracted with hot MeOH (50 °C, 3 × 200 mL). The resulting MeOH extracts (800 mL) were combined, filtered under reduced pressure over a short path of sea sand, and the filtrate evaporated to dryness in vacuo. The resulting residue was subjected to column chromatography on Si gel (60 g) using hexane–EtOAc 3:1 as solvent to give (+)-sesamolol (**1**) (20 mg) and (+)-sesamin (**5**) (10 mg).⁹ Further elution with hexane–EtOAc 1:1 gave two fractions; HPLC–MS of the latter fraction gave a single component at the same retention time as **2** with identical UV and MS spectra to those of the synthetic (+)-episesaminone (**2**).

Hydrolysis of **1** to give the known (+)-samin **7**,¹³ subsequent conversion to (+)-acuminatolide (**8**), and complete physical and spectroscopic data of **7** and **8** are described elsewhere.¹

Dimer 6: IR (CHCl₃) ν_{\max} 1503, 1443 (C=C_{Ar}), 1248 (C–O) cm⁻¹; UV (CHCl₃) λ_{\max} (log ϵ) 243 (3.8), 288 (3.9), 337 (1.32) nm; mp 188–189 °C (MeOH), (lit.¹⁴ 191–192 °C); [α]_D +160.1° (*c* 1.37, CHCl₃) (lit.¹⁴ [α]_D +143° (*c* 1.74, CHCl₃)); MS *m/z* 482 [M⁺] (5), 233 [C₁₃H₁₃O₄⁺] (10), 203 [ArC₅H₆O⁺] (15), 189 (5), 163 (10), 150 [ArCHO⁺] (95), 149 [ArCO⁺] (100), 135 [ArCH₂⁺] (90), 121 [Ar⁺] (20); HRMS 482.1581 (calcd for C₂₆H₂₆O₉, 482.1517); ¹H-NMR δ 6.90–6.75 (m, 6H, H-2, H-5 and H-6), 5.96 (s, 4H, OCH₂O), 5.29 (s, 2H, H-7'), 4.37 (d, *J* = 8.4 Hz, 2H, H-7), 4.38 (dd, *J* = 9.2 Hz, *J* = 7.8 Hz, 2H) and 4.05–3.95 (m, 4H) and 3.58 (dd, *J* = 9.2 Hz, *J* = 7.8 Hz, 2H, H-9 and H-9'), 3.05–2.95 (m, 2H, H-8'), 2.90–2.85 (m, 2H, H-8); ¹³C-NMR (DEPT mode) δ 148.06 (s, C-3), 147.39 (s, C-4), 134.63 (d, C-1), 119.78 (d, C-6), 108.25 (d, C-5), 106.65 (d, C-2), 102.96 (d, C-7), 101.17 (t, OCH₂O), 87.06 (d, C-7), 71.44 (t, C-9), 69.43 (t, C-9'), 52.84, 52.77 (d, C-8 and C-8') ppm.

Synthesis of (+)-Episesaminone (2). A suspension of magnesium turnings (5.2 mg, 1.2 equiv) and 1-bromo-3,4-(methylenedioxy)benzene (1.1 equiv, 23.5 mL) in dry THF (5 mL) was heated under argon until reflux began, this being maintained for 3 h. The resulting suspension was cooled to room temperature and added dropwise to a solution of **8** (44 mg, 0.177 mmol) in dry THF (50 mL). After being stirred for 12 h, the solvent was removed in vacuo, with the resulting residue subjected to column chromatography (hexane–EtOAc, 3:1 then 1:1) to give **2** (18.4 mg, yield 29%, conversion 47% based on starting material) and unreacted starting material **8** (18 mg). The [α]_D (120°; *c* 0.127, CHCl₃), MS, HRMS, ¹H and ¹³C NMR, IR, and UV spectra were identical with naturally occurring (+)-episesaminone (**2**) except for the optical rotation value, i.e., [α]_D +120° (*c* 0.127, CHCl₃) versus that of the natural product **2** {[α]_D +126.2° (*c* 0.127, CHCl₃) see above}.

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References and Notes

- (1) Marchand, P. A.; Zajicek, J., Lewis, N. G. *Can. J. Chem.* **1997**, *75*, 840–849.
- (2) Davin, L. B.; Wang, H.-B.; Crowell, A. L.; Bedgar, D. L.; Martin, D. M.; Sarkanen S.; Lewis, N. G. *Science* **1997**, *275*, 362–366.
- (3) Dinkova-Kostova, A. T.; Gang, D. R.; Davin, L. B.; Bedgar, D. L.; Chu, A.; Lewis, N. G. *J. Biol. Chem.* **1996**, *271*, 29473–29482.
- (4) Chu, A.; Dinkova, A.; Davin, L. B.; Bedgar D. L.; Lewis, N. G. *J. Biol. Chem.* **1993**, *268*, 27026–27033.
- (5) Gang, D. R.; Dinkova-Kostova, A. T.; Davin, L. B.; Lewis, N. G. *ACS Symp. Ser.* **1997**, *658*, 58–89.
- (6) Davin, L. B.; Lewis, N. G. *ACS Symp. Ser.* **1994**, *562*, 202–246.
- (7) Pelter, A.; Ward, S.; Rao, E. V.; Sastry, K. V. *Tetrahedron* **1976**, *32*, 2783–2788.
- (8) Kinjo, J.; Higuchi, H.; Fukui, K.; Nohara, T. *Chem. Pharm. Bull.* **1991**, *39*, 2952–2955.
- (9) Banerji, A.; Sarkar, M.; Ghosal, T.; Pal, S. C.; Shoolery, J. N. *Tetrahedron* **1984**, *40*, 5047–5052.
- (10) Rao, K. V.; Wu, W.-N. *Lloydia* **1978**, *41*, 56–62.
- (11) Chiung, Y.-M.; Hayashi, H.; Matsumoto, H.; Otani, T.; Yoshida, K.-I.; Huang, M.-Y.; Chen, R.-X.; Liu, J.-R.; Nakayama, M. *J. Antibiot.* **1994**, *47*, 487–491.
- (12) Takano, S.; Ohkawa, T.; Tamori, S.; Satoh, S.; Ogasawara, K. *J. Chem. Soc., Chem. Commun.* **1988**, 189–191.
- (13) Davenport, J. B.; Sutherland, M. D. *Aust. J. Chem.* **1954**, *7*, 384–386.
- (14) Haslam, E.; Haworth, R. D. *J. Chem. Soc.* **1955**, 827–833.
- (15) Fukuda, Y.; Isobe, M.; Nagata, M.; Osawa, T.; Namiki, M. *Heterocycles* **1986**, *24*, 923–926.
- (16) Corey, E. J.; Su, W.-G. *J. Am. Chem. Soc.* **1987**, *109*, 7534–7536.
- (17) Jakupovic, J.; Pathak, V. P.; Bohlmann, F.; King, R. M.; Robinson, H. *Phytochemistry* **1987**, *26*, 803–807.
- (18) Ishibashi, H.; Su So, T.; Okochi, K.; Sato, T.; Nakamura, N.; Nakatani, H.; Ikeda, M. *J. Org. Chem.* **1991**, *56*, 95–102.
- (19) Adriani, W. Z. *Unters. Lebensmittel* **1928**, *56*, 187–194.
- (20) Iida, T.; Nakano, M.; Ito, K. *Phytochemistry* **1982**, *21*, 673–675.
- (21) Brieskorn, C. H.; Huber, H. *Tetrahedron Lett.* **1976**, *26*, 2221–2224.
- (22) Osawa, T.; Nagata, M.; Namiki, M.; Fukuda, Y. *Agric. Biol. Chem.* **1985**, *49*, 3351–3352.
- (23) Fonseca, S. F.; Nielsen, L. T.; Ruveda, E. A. *Phytochemistry* **1979**, *18*, 1703–1708.

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